

## **Thymidine incorporation assay with Balb/c 3T3 cells (96-well cluster plate)**

### **Material:**

- 1% Gelatine in ddH<sub>2</sub>O, PBS
- Growth medium: DMEM (4,5g Glucose), 1% Pen/Strep, 1% Glutamin, 10% Calf Serum (HyClone: Cat.No. SH30072.03)
- Basal medium: DMEM, 1% Pen/Strep, 1% Glutamin, 2,5% CS
- <sup>3</sup>H-Thymidine 1mCi/ml, 1:40 verdünnt in PBS (0,025mCi/ml)
- PBS, Methanol, 5% TCA, ddH<sub>2</sub>O, 0,3M NaOH (use at 4°C!)

### **Protocol:**

- Expand Balb/c 3T3 cells in growth medium (10% CS) and use cells at a confluence of about 70-90%
- coat a 96-well cluster plate with gelatine (1% in water) and incubate for 15 min at 37°C  
    ↯ fill at least the outer row with PBS because of evaporation over the long time
- plate cells with a density at  $2 \times 10^3$  cells/well in basal medium (2.5% CS)
- incubate cells 7 to 14 d at 37°C and 5% CO<sub>2</sub>
- stimulate the cells with the desired growth factors (as positive control 10µl CS is used)
- add 10?l <sup>3</sup>H-Thymidine solution [0.025mCi/ml] per well (=0.25?Ci)
- incubate cells 36 to 64 h
- carefully remove the medium
- Washing steps: (250?l/well)

PBS	1x
MeOH	2x 5min
TCA	2x 10min
H <sub>2</sub> O	1x
- lyse cells in 250?l 0.3M NaOH per well
- transfer 2.5 ml ECO Plus into the appropriate scintillation vials
- transfer cell lysate into the scintillation vials
- count by liquid scintillation (β-counter; Beckmann Instruments)